

New apocarotenoids and β -carotene cleavage in *Blakeslea trispora*†Alejandro F. Barrero,*^a M. Mar Herrador,^a Pilar Arteaga,^a Jesús Gil,^a Jose-Antonio González,^a Eugenio Alcalde^b and Enrique Cerdá-Olmedo*^b

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Mixed cultures of strains of opposite sex (“mated” cultures) of *Blakeslea trispora* contain trisporic acids and other apocarotenoids, some of which mediate the sexual responses of this fungus and other Mucorales. In mated cultures of the wild-type strains F986 and F921 we identified eleven apocarotenoids: two C₁₈ trisporoids, three C₁₅ compounds with a monocyclofarnesane skeleton, a C₁₃ compound, and five C₇ compounds with a 2-methylhexane skeleton. Six of them are new natural products and two others are new for *Blakeslea*. Their structures were established by NMR and mass spectra and those of the C₇ and C₁₃ compounds were confirmed by chemical synthesis. The finding of these compounds and the presence of approximately equimolar amounts of the C₁₈, C₁₅, and C₇ families led to the conclusion that β -carotene is initially split in three fragments by cleavage of its 13,14 and 11',12' double bonds.

Introduction

β -Carotene (**1**, Fig. 1) is a natural pigment, antioxidant, and pro-vitamin A with many applications in the alimentary, pharmaceutical, and cosmetic industries.¹ It is obtained commercially by either chemical synthesis or biotechnology, particularly from the fungus *Blakeslea trispora* (syn. *Choanephora trispora*, Mucoromycotina, Mucorales, Choanephoraceae). The wild-type strains of this fungus belong to either the (+) or the (–) sex, and many pairs of strains of opposite sex, cultured together (“mated” cultures), increase their β -carotene content and start the morphological program of the sexual cycle. These physiological effects were attributed to apocarotenoids such as trisporic acid **2** and similar compounds present in mated cultures of *Blakeslea*.² The culture media of *Blakeslea* contain apocarotenoids with 18 carbons (**2–14**),^{2a,3} called trisporoids, or with 15 carbons (**15–17**),⁴ often called apotrisporoids because they were presumed to derive from the former.⁵ On the other hand two apocarotenoids with 7 carbons (**23**, **24**) have been found recently in cultures from another Mucoral, *Phycomyces blakesleeanus*.^{6a}

We have studied the apocarotenoids in cultures of our standard wild types of *B. trispora*. The structure of the identified products and their relative amounts indicate that the apocarotenoids of *B. trispora* derive from an asymmetrical double cleavage of β -carotene.

Results and discussion

Results

We analyzed the agar medium where the strains F986, sexually (+), and F921, sexually (–) of *B. trispora* had been cultured together for three days. A clean medium, obtained by freezing and squeezing the agar and centrifuging the resulting liquid, was brought to pH 8 with NaOH and extracted with ethyl acetate. This “neutral extract” was fractionated by semi-preparative normal-phase HPLC. The remaining water solution was brought to pH 2 with HCl and extracted with ethyl acetate. This “acid extract” was fractionated in the same way after methylation with (trimethylsilyl)diazomethane (TMSCHN₂).

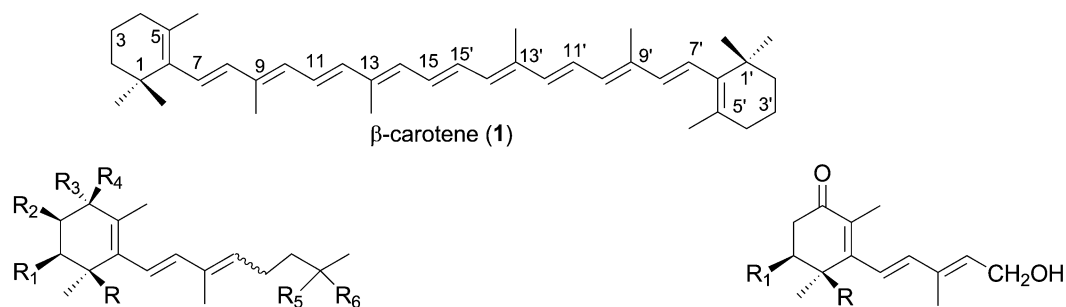
We isolated eleven apocarotenoids: three C₁₅ (**18** and **19**, as methyl esters, and **17**), two C₁₈ (the 9*E* and 9*Z* isomers of **2**, both as methyl esters), five C₇ (**20–24**), and one C₁₃ apocarotenoid (**25**, as methyl ester) (Fig. 2). The structures of five of them (**17**, **23**, **24**, and the 9*E* and 9*Z* isomers of **2**) were determined by comparing their spectroscopic data with those reported in the literature.^{4c,6} Apocarotenoid **25** is a new natural product, but its methyl ester (**25a**) has been described as an intermediate in the synthesis of trisporic acid B.⁷ The five remaining apocarotenoids (**18–22**) are new to science.

Apocarotenoid **22** is the most abundant C₇ apocarotenoid in our cultures. Its molecular formula C₇H₁₂O₂ was deduced from HRFABMS. Its IR spectrum showed the absorption bands of a hydroxyl group at 3447 cm^{–1} and of a conjugated diene at 1600 and 1650 cm^{–1}. The ¹³C NMR spectrum showed seven signals: two primary alcohol signals at δ 63.6 and 68.3, a methyl signal at δ 14.1, and four signals for disubstituted and trisubstituted double bonds at δ 137.9 (C), 131.8 (CH), 127.1 (CH) and 123.8 (CH). The 2-methyl-2,4-hexadiene skeleton was established by the direct

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Trisporic acid C (2) R = COOH R₁ = R₂ = H R₃, R₄ = O R₅ = H R₆ = OH
 Trisporic acid A (3) R = COOH R₁ = R₂ = R₅ = R₆ = H R₃, R₄ = O
 Trisporic acid B (4) R = COOH R₁ = R₂ = H R₃, R₄ = R₅, R₆ = O
 Trisporic acid D (5) R = COOH R₁ = OH R₂ = H R₃, R₄ = R₅, R₆ = O
 Trisporic acid E (6) R = COOH R₁ = H R₂ = OH R₃, R₄ = O R₅ = H R₆ = OH
 Trisporin B (7) R = CH₃ R₁ = R₂ = H R₃, R₄ = R₅, R₆ = O
 Trisporin C (8) R = CH₃ R₁ = R₂ = H R₃, R₄ = O R₅ = H R₆ = OH
 Trisporol B (9) R = CH₂OH R₁ = R₂ = H R₃, R₄ = R₅, R₆ = O
 Trisporol C (10) R = CH₂OH R₁ = R₂ = H R₃, R₄ = O R₅ = H R₆ = OH
 Methyl 4-dihydrotrisporate B (11) R = COOMe R₁ = R₂ = H R₃ = H R₄ = OH R₅, R₆ = O
 Methyl 4-dihydrotrisporate C (12) R = COOMe R₁ = R₂ = H R₃ = H R₄ = OH R₅ = H R₆ = OH
 Methyl trisporate B (13) R = COOMe R₁ = R₂ = H R₃, R₄ = R₅, R₆ = O
 Methyl trisporate C (14) R = COOMe R₁ = R₂ = H R₃, R₄ = O R₅ = H R₆ = OH

Apotrisporin (15) R = CH₃ R₁ = H
 Apotrisporin E (16) R = CH₃ R₁ = OH
 Apotrisporol (17) R = CH₂OH R₁ = H

Fig. 1 Apocarotenoids isolated from cultures of different strains of *Blakeslea trispora*.

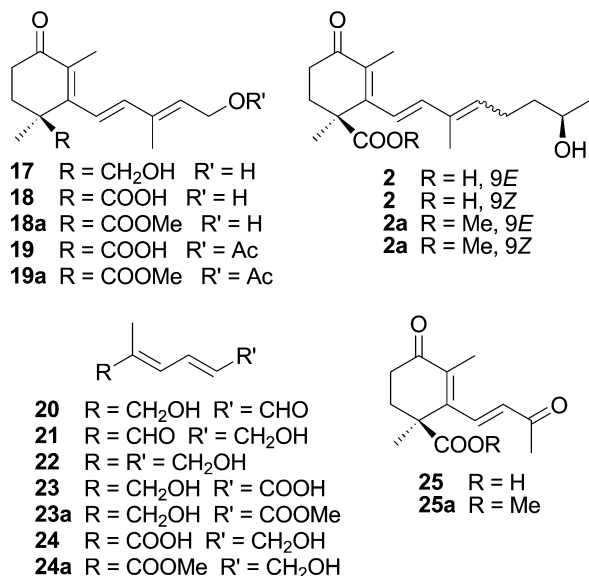


Fig. 2 Apocarotenoids isolated from mated cultures of the *B. trispora* wild-type strains F986 and F921.

coupling of the three olefinic protons in the ¹H NMR spectrum at δ 6.43 (dd, *J*₁ = 11.0 Hz, *J*₂ = 15.1 Hz), 6.02 (d, *J* = 11.0 Hz), and 5.78 (dt, *J*₁ = 5.5 Hz, *J*₂ = 15.1 Hz). The *E* stereochemistry of the disubstituted double bond was deduced from its coupling constant (15.1 Hz) and that of the trisubstituted one, from the chemical shifts at C-1 (δ 68.3) and C-7 (δ 14.1) in the ¹³C NMR spectrum.⁸ We conclude that compound **22** is (2*E*,4*E*)-2-methyl-2,4-hexadiene-1,6-diol.

Compound **20** and **21** shared most of their ¹H NMR signals with compound **22** (Table 1). The ¹H NMR spectrum of compound **20** lacked one of the CH₂OH signals (δ 4.15, d, *J* = 5.5 Hz) of compound **22**, but showed a CHO signal (δ 9.62, d, *J* = 8.0 Hz). The ¹H

Table 1 ¹H and ¹³C NMR data of **20–22**^a

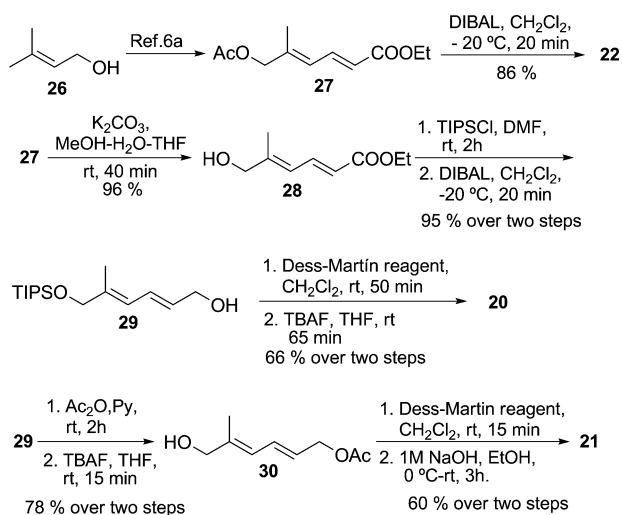
20	21		22		
	δ _H	δ _C	δ _H	δ _C	
1	9.62 d (8.0)	194.1	9.48 s	68.3	
2	6.09 dd (8.0; 15.1)	121.7	—	137.9	
3	7.65 dd (11.6; 15.1)	148.7	7.03 d (11.2)	6.02 d (11.0)	123.8
4	6.48 d (11.6)	131.7	6.85 dd (11.2, 15.1)	6.43 dd (11.0; 15.1)	131.8
5	—	152.6	6.44 dt (4.6; 15.1)	5.78 dt (5.5; 15.1)	127.1
6	4.18 br s	67.0	4.31 d (4.6)	4.15 d (5.5)	63.6
7	1.92 s	14.6	1.82	1.73	14.1

^a δ in ppm. *J* in Hz in parentheses.

NMR spectrum of compound **21** lacked the other CH₂OH signal (δ 4.02, s) of compound **22**, but showed a CHO signal (δ 9.48, s).

The proposed structures for **20–22** were confirmed by chemical synthesis (Scheme 1).

An efficient synthesis of acetoxyester **27** from the commercial prenol **26** (3-methylbut-2-en-1-ol) has been reported.^{6a} Reduction of **27** with excess diisobutylaluminium hydride (DIBAL) produced **22** with a yield of 86%, whereas the chemoselective saponification of the acetoxy group with K₂CO₃ produced the hydroxyester **28** with a yield of 96%. Protection of **28** with triisopropylsilyl chloride (TIPSCl) and reduction with DIBAL produced to **29** with a yield of 95%. Oxidation of **29** with the Dess–Martin reagent and removal of the silyl protection with tetrabutylammonium fluoride (TBAF) produced **20** with a yield of 66%. Acetylation of **29** with Ac₂O/pyridine and removal of the silyl protection with TBAF produced **30** with a yield of 78%. Oxidation of **30** with the Dess–Martin reagent and saponification with 1 M NaOH produced **21** with a yield of 60%. The spectroscopic data of the synthetic products coincided with those of the natural products.



Scheme 1 Synthesis of the apocarotenoids 20–22.

Table 2 ¹H and ¹³C NMR data of 17, 18a, 19a^a

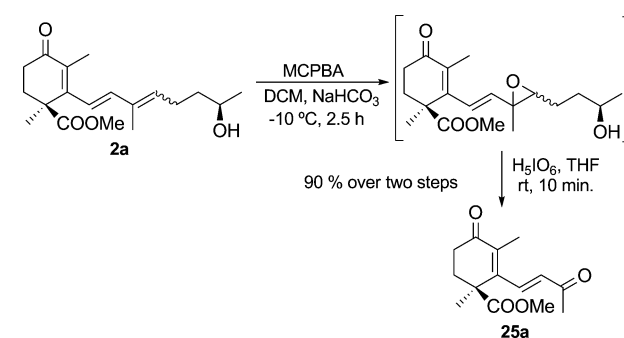
C	17		18a		19a	
	δ_{H}	δ_{C}	δ_{H}	δ_{C}	δ_{H}	δ_{C}
1	—	41.2	—	46.9	—	47.0
2a	1.69–1.73 m	33.9	1.92–1.97 m	34.5	1.91–1.96 m	34.5
2b	2.23–2.29 m	—	2.37–2.43 m	—	2.36–2.41 m	—
3a	2.53–2.60 m	31.8	2.50–2.54 m	33.5	2.45–2.53 m	33.6
3b	2.53–2.60 m	—	2.50–2.54 m	—	2.45–2.53 m	—
4	—	199.0	—	197.4	—	197.8
5	—	135.5	—	132.5	—	133.0
6	—	157.4	—	152.3	—	152.2
7	6.22 d (16.3)	124.4	6.34 s	124.6	6.30 d (16.5)	125.2
8	6.26 d (16.3)	140.3	6.34 s	139.8	6.35 d (16.5)	139.1
9	—	135.5	—	135.9	—	137.9
10	5.76 t (6.8)	132.8	5.77 dd (1.1; 6.6)	134.1	5.67 br t (6.9)	128.7
11	4.34 d (6.8)	59.4	4.33 d (6.6)	59.6	4.72 d (6.9)	61.2
12	1.86	13.8	1.83 s	12.5	1.85 s	12.6
13a	3.44 d (11.0)	69.5	—	176.3	—	176.3
13b	3.73 d (11.0)	—	—	—	—	—
14	1.14	21.7	1.54 s	23.0	1.49 s	23.1
15	1.87	12.5	1.94 s	12.3	1.92 s	12.5
OH	1.53 br s	—	—	—	—	—
OMe	—	—	3.69 s	52.5	3.67 s	52.6
Ac	—	—	—	—	—	170.9
Ac	—	—	—	—	2.06 s	21.0

^a δ in ppm. *J* in Hz in parentheses.

The molecular formula of methyl ester of apocarotenoid **18** (**18a**) was C₁₆H₂₂O₄ according to its HRFABMS. Its structure was established from its NMR data (Table 2), which indicated a COOMe group attached to C1, instead of the CH₂OH group of the well known apotrisporol (**17**).

The spectroscopic data of methyl ester of **19** (**19a**) led us to the conclusion that **19a** was the acetyl derivative of **18a** (Table 2).

Compound **25** is the first C₁₃ apocarotenoid isolated in the Mucorales. It is found in very small amounts in the cultures and could have been formed from a C₁₅ or C₁₈ apocarotenoid by spontaneous oxidative breakage or a retroaldol reaction. It was identified by comparison of spectral data of its methyl ester **25a** with those published for a synthetic compound.⁷ The structure was confirmed by semisynthesis (Scheme 2).

Scheme 2 Synthesis of apocarotenoid **25a**.

The reaction of the methyl ester of the trisporic acid **C** (**2a**) with *m*-chloroperoxybenzoic acid (MCPBA) produced the regioselective epoxidation of the 9,10 double bond. Treatment of the crude product with periodic acid produced **25a** with an overall yield of 90%. Its spectroscopic data coincided with those of the methyl ester of the natural product.

A semiquantitative analysis based on the ¹H NMR signals from spectra of the “neutral extract” and “acid extract” indicated that the three groups of apocarotenoids (trisporoids with 18 carbons, cyclofarnesoids with 15 carbons, and methylhexadienes with 7 carbons) were found in approximately equimolar amounts. For this summation the small amount of the only C₁₃ apocarotenoid can be disregarded.

Small amounts of compounds **23** and **24** and of three other compounds were detected by their UV absorption in the chromatograms of the wild type F921, but not in those of strain SB64, a mutant derived from it and completely devoid of β -carotene. This confirms that **23** and **24** are apocarotenoids, as shown already for *Phycomyces*.^{6a}

Discussion

There are four reasons to think that the compounds described here derive from β -carotene: the structural similarity between them and three segments of the β -carotene molecule (Fig. 3); the approximate equimolar amounts of trisporoids, cyclofarnesoids, and methylhexadienes; the early label experiments by Austin *et al.*,^{2b} and the comparisons^{6a} of wild-type strains with mutants devoid of β -carotene.

These arguments indicate that the three families of apocarotenoids of *Blakeslea* result from the double oxidative cleavage of β -carotene at its 13,14 and 11',12' double bonds. This mechanism was already shown for *Phycomyces*^{6a} and is likely to be common to all Mucorales.

Our results with *Blakeslea* and the previous ones with *Phycomyces* argue strongly against the hypothesis^{2b} that trisporic acids derive from β -carotene *via* retinal. This hypothesis was already suspicious because of the failure of the efforts to identify retinal in *Phycomyces* cultures. Our results also reject the alternative hypothesis that β -carotene is cut twice at its 13,14 and 13',14' double bonds to produce the first trisporoid and that C₁₅ apocarotenoids result from the secondary loss of three carbon atoms of the side chain.

The two C₇ compounds with an aldehyde group and a hydroxy group **20** and **21** were the least abundant and probably represent metabolic intermediates. The likely biosynthetic pathway would

ester mixture (80 mg) that was fractionated by semi-preparative HPLC. The fraction (10.5 < RT < 13.1 min, 3.4 mg) contained **19a**. The fraction (13.4 < RT < 15.9 min, 9 mg) contained a 2 : 1 mixture of **23a** and **24a**. The fraction (16.1 < RT < 16.5 min, 9.5 mg) contained **2a(9E)**. The fraction (16.5 < RT < 17.0 min, 3.9 mg) contained a 2 : 2 : 1 mixture of **2a(9E)**, **2a(9Z)** and **25a**. The fraction (17.0 < RT < 18.4 min, 11.9 mg) contained a 5 : 1 mixture of **2a(9Z)** and **25a**. The fraction (18.4 < RT < 20.7 min, 10 mg) contained **2a(9Z)**. The fraction (20.7 < RT < 23.1 min, 4 mg) contained a 2 : 1 mixture of **2a(9Z)** and **18a**. The fraction (23.1 < RT < 25.6 min, 6 mg) contained **18a**.

(2E,4E)-6-hydroxy-5-methylhexa-2,4-dienoic acid (23, as methyl ester) and (2E,4E)-6-hydroxy-2-methylhexa-2,4-dienoic acid (24, as methyl ester). Their spectroscopic data coincided with those previously reported.^{6a}

Apotrisporic acid (18, as methyl ester). Colourless syrup. ¹H NMR (500 MHz, CDCl₃): see Table 2; ¹³C NMR (125 MHz, CDCl₃): see Table 2; *m/z* (HRMS(FAB)) 301.1414 (M + Na). C₁₆H₂₂O₄Na requires 301.1416).

Apotrisporic acid 11-acetate (19, as methyl ester). Colourless syrup. ¹H NMR (500 MHz, CDCl₃): see Table 2; ¹³C NMR (125 MHz, CDCl₃): see Table 2; *m/z* (HRMS(FAB)) 343.1518 (M + Na). C₁₈H₂₄O₅Na requires 343.1521).

(7E,9E)-Trisporic acid C (2(9E)), as methyl ester. Colourless syrup. Its spectroscopic data coincided with those previously reported.^{6c}

(7E,9Z)-Trisporic acid C (2(9Z)), as methyl ester. Colourless syrup. Its spectroscopic data coincided with those previously reported.^{6b,c}

Methyl (E)-1,3-dimethyl-4-oxo-2-(3-oxo-1-butenyl)-2-cyclohexene-1-carboxylate (25a). Colourless syrup. Its spectroscopic data coincided with those previously reported.⁷

Preparation of 25a

m-CPBA (64 mg, 2.27 mmol) in DCM (0.8 mL) was added dropwise to a mixture of **7** (50 mg, 0.16 mmol) in dichloromethane (DCM) (2.6 mL) and 0.5 M NaHCO₃ (0.5 mL) under stirring and argon at 0 °C. After 2.5 h, the mixture was diluted with *t*-butyl methyl ether and washed with 0.5 M NaHCO₃, a saturated solution of sodium thiosulfate and brine, successively. The organic phase was dried over anhydrous Na₂SO₄ and filtered. The residue obtained (50 mg) after removing the solvent under low pressure was dissolved in THF (2.6 mL) and H₃IO₆ (35.7 mg, 0.16 mmol) was added at room temperature. After 10 min, water (10 mL) was added to the mixture and tetrahydrofuran (THF) was removed under low pressure. The aqueous phase was extracted with *t*-butyl methyl ether and the organic phase was dried over anhydrous Na₂SO₄ and filtered. The residue obtained after removing the solvent under low pressure was chromatographed in a silica gel column (hexane : *t*-butyl methyl ether, 1/1, v/v) to obtain **25** (36 mg, 90%).

Preparation of (2E,4E)-6-hydroxy-5-methyl-2,4-hexadienal (20)

Saponification of 27. Preparation of ethyl (2E,4E)-6-hydroxy-5-methyl-2,4-hexadienoate (28). K₂CO₃ (20 g) and K₂CO₃ 2 M

(43 mL) were added to a solution of **27** (1.4 g, 6.65 mmol) in THF (84 mL) and MeOH (25 mL). The mixture was stirred for 40 min at room temperature. Then the solvent was evaporated under low pressure to obtain a residue that was extracted with Et₂O. The organic phase was washed with brine, dried over anhydrous Na₂SO₄ and filtered. The residue obtained after removing the solvent under low pressure was chromatographed in a silica gel column (petroleum ether (bp 30–40 °C)/Et₂O, 50/50, v/v) to obtain **28** (1.08 g, 96%).

Ethyl (2E,4E)-6-hydroxy-5-methyl-2,4-hexadienoate (28). Yellow oil. IR ν_{\max} (film)/cm⁻¹ 3435, 1641, 1308, 1276, 1160, 1035 and 983; δ_{H} (300 MHz; CDCl₃; Me₄Si) 7.62 (1H, dd, *J* 11.7 and 15.2, H-3), 6.29 (1H, d, *J* 11.7, H-4), 5.91 (1H, d, *J* 15.2, H-2), 4.24 (2H, q, *J* 7.1, OCH₂CH₃), 4.19 (2H, s, H-6), 1.92 (3H, s, H-7), 1.72 (1H, br s, OH) and 1.33 (3H, t, *J* 7.1, OCH₂CH₃); δ_{C} (75 MHz; CDCl₃; Me₄Si) 167.5 (C, C-1), 147.2 (C, C-5), 140.0 (CH, C-3), 121.8 (CH, C-4), 121.0 (CH, C-2), 67.6 (CH₂, C-6), 60.4 (CH₂, OCH₂CH₃), 14.7 (CH₃, OCH₂CH₃)^a and 14.4 (CH₃, C-7)^a (^aSignals with the same letter are exchangeable); *m/z* (HRMS(FAB)) 193.0844 (M + Na). C₉H₁₄O₃Na requires 193.0841).

Preparation of hydroxy-ether 29. Imidazole (1.07 g, 15.73 mmol) and TIPSCl (1.45 g, 7.42 mmol) were added to a solution of **28** (1.07 g, 6.29 mmol) in dry DMF (1.1 mL) under argon and stirring at room temperature. After 2 h, the mixture was fractionated in *t*-BuOMe:H₂O (2:1). The organic phase was washed successively with HCl 2 N and brine and dried over anhydrous Na₂SO₄. Once the solvent was evaporated under low pressure we obtained a crude product (2.14 g), which was dissolved in dry DCM (121 mL) under argon and chilled at –20 °C. After adding DIBAL (1 M in hexane, 16 mL) the solution was stirred for 20 min and then, H₂O (49 mL) was added. The mixture was stirred at room temperature for 20 min and filtered through a layer of silica gel/anhydrous Na₂SO₄ (2/1, w/w). The layer was washed with Et₂O and the organic phase was evaporated under low pressure to afford a crude product, which was chromatographed over silica gel column (petroleum ether (bp 30–40 °C)/Et₂O, 85/15, v/v) to obtain **29** (1.7 g, 95%).

(2E,4E)-6-Triisopropylsilyloxy-5-methyl-2,4-hexadien-1-ol (29). Colourless syrup. IR ν_{\max} (film)/cm⁻¹ 3327, 2942, 2865, 1641, 1462, 1385, 1113, 1085, 1065, 995 and 882; δ_{H} (500 MHz; CDCl₃; Me₄Si) 6.50 (1H, dd, *J* 11.2 and 15.1, H-3), 6.15 (1H, d, *J* 11.2, H-4), 5.80 (1H, dt, *J* 6.0 and 15.1, H-2), 4.20 (2H, d, *J* 6.0, H-1), 4.14 (2H, s, H-6), 1.72 (3H, s, H-7) and 1.04–1.18 (21H, m, TIPS); δ_{C} (125 MHz; CDCl₃; Me₄Si) 138.3 (C, C-5), 130.8 (CH, C-3), 127.7 (CH, C-2), 122.2 (CH, C-4), 67.9 (CH₂, C-6), 63.8 (CH₂, C-1), 18.1 (6CH₃, TIPS), 13.9 (CH₃, C-7) and 12.1 (3CH, TIPS); *m/z* (HRMS(FAB)) 307.2071 (M + Na). C₁₆H₃₂O₂Na requires 307.2070).

Preparation of hydroxy-aldehyde 20. The Dess–Martin reagent (1.01 g) was added to a solution of **29** (800 mg, 2.82 mmol) in dry DCM (31 mL) at room temperature under argon. The mixture was left for 50 min under stirring. A saturated solution of Na₂S₂O₃ and NaHCO₃ was then added dropwise to the mixture and extracted with Et₂O. The organic phase was washed with brine, dried over anhydrous Na₂SO₄, and filtered. The residue obtained after evaporating the solvent under low pressure was dissolved in petroleum ether (bp 30–40 °C)/Et₂O (3/1, v/v) and

filtered through silica gel. The residue (713 mg) obtained after evaporating the solvent under low pressure was dissolved in dry THF (37 mL) and mixed with TBAF (1 M in THF, 16 mL), stirred at room temperature for 65 min, diluted with Et₂O (15 mL), and washed with brine. The organic phase was dried over anhydrous Na₂SO₄ and filtered. The residue obtained after removing the solvent under low pressure was chromatographed in a silica gel column (petroleum ether (bp 30–40 °C)/Et₂O, 35/65, v/v) to obtain **20** (234 mg, 66%).

Preparation of (2E,4E)-6-hydroxy-2-methyl-2,4-hexadienal (**21**)

Preparation of hydroxy-acetate 30. A mixture of **29** (730 mg, 2.57 mmol), dry pyridine (11 mL) and acetic anhydride (0.5 mL) was left at room temperature for 2 h and then was worked up as usual to give a crude product (835 mg), which was dissolved in dry THF (33 mL), mixed with TBAF (1 M in THF, 2.2 mL), stirred at room temperature under argon for 15 min, diluted with Et₂O (15 mL), and washed with brine. The organic phase was dried over anhydrous Na₂SO₄ and filtered. The residue obtained after removing the solvent under low pressure was chromatographed in a silica gel column (petroleum ether (bp 30–40 °C)/Et₂O, 1/1, v/v) to obtain **30** (340 mg, 78%).

(2E,4E)-6-Hydroxy-5-methyl-2,4-hexadienyl acetate (30). Colourless syrup. IR ν_{\max} (film)/cm⁻¹ 3420, 2917, 2861, 1737, 1662, 1445, 1380, 1365, 1235, 1113, 1071, 1023 and 971; δ_{H} (400 MHz; CDCl₃; Me₄Si) 6.51 (1H, dd, *J* 11.0 and 15.2, H-3), 6.05 (1H, d, *J* 11.0, H-4), 5.71 (1H, dt, *J* 6.3, 15.2, H-2), 4.58 (2H, d, *J* 6.3, H-1), 4.03 (2H, s, H-6), 2.06 (1H, br s, OH), 2.03 (3H, s, COCH₃) and 1.75 (3H, s, H-7); δ_{C} (100 MHz; CDCl₃; Me₄Si) 171.9 (C, COCH₃), 140.0 (C, C-5), 131.0 (CH, C-3), 126.8 (CH, C-2), 123.9 (CH, C-4), 68.3 (CH₂, C-6), 65.4 (CH₂, C-1), 21.1 (CH₃, COCH₃) and 14.2 (CH₃, C-7); *m/z* (HRMS(FAB)) 193.0838 (M + Na. C₉H₁₄O₃Na requires 193.0841).

Preparation of hydroxy-aldehyde 21. The Dess–Martin reagent (1.05 g) was added to a solution of **30** (340 mg, 2.00 mmol) in dry DCM (11 mL) at room temperature under argon. The mixture was left for 15 min under stirring. A saturated solution of Na₂S₂O₃ and NaHCO₃ was then added dropwise to the mixture and extracted with Et₂O. The organic phase was washed with brine, dried over anhydrous Na₂SO₄, and filtered. The residue obtained after evaporating the solvent under low pressure was dissolved in petroleum ether (bp 30–40 °C)/Et₂O (3/1, v/v) and filtered through silica gel. The residue (245 mg) obtained after evaporating the solvent under low pressure was dissolved in EtOH (7.60 mL) and NaOH 1 M (3.80 mL) was added dropwise at 0 °C. The mixture was then left at room temperature for 3 h. The solution was neutralized with HCl 1 N (3.80 mL) and the solvent was evaporated under low pressure. The residue was extracted with EtOAc and the solvent was evaporated under low pressure to obtain **21** (151 mg, 60%).

Preparation of (2E,4E)-2-methyl-2,4-hexadiene-1,6-diol (**22**)

DIBAL (1 M in hexane, 2 mL) was added to a cold (–20 °C) solution of **30** (100 mg, 0.78 mmol) in dry CH₂Cl₂ (9 mL) under argon with stirring. After 20 min, water (3 mL) was added to the mixture, and it was stirred at room temperature for 20 min. The mixture was filtered through a layer of silica gel/anhydrous Na₂SO₄ (2/1, w/w). The layer was washed with Et₂O and the organic phase was evaporated under low pressure to afford a crude product, which was chromatographed over silica gel column (petroleum ether (bp 30–40 °C)/Et₂O, 35/65, v/v) to obtain **22** (65 mg, 86%).

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